Characterization and evaluation of *Berberis microphylla* G. Forst pollen grains

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Abstract: *Berberis microphylla*, commonly known as “calafate”, is a non-timber forest product native from Patagonia, and its berries possess highlighted nutraceutical value. The objective of this research was to describe the morphology and anatomy of pollen grains of *Berberis microphylla* G. Forst genotypes growing spontaneously on the island of Tierra del Fuego (Argentina), and evaluate their vitality and germination. Pollen grain diameter varied from 40 to 47.26 μm, the pollen grains of 124 and 201 genotypes being significantly smaller than the others. Vitality measured by DAPI methodology was also variable among genotypes, although always about 50%. *In vitro* germination of pollen grains measured one day after the flowers were collected was very high for some genotypes (near 80%), and then decreased after 21 days of storage, except for genotype 123 whose germination value increased from 44.34 to 69%. The significant variability found in pollen performance (size, viability and germination) among *B. microphylla* genotypes from a natural population could be interpreted as an enhanced survival strategy to maximize reproduction fitness, with a marked capacity of response to environmental changes. High viable pollen frequency together with germination percentages observed in all the genotypes tested could indicate a good fertilization process. The correlation observed between size and germination percentage could be used as markers of pollen grain performance, paving the way for possible *B. microphylla* breeding.

1. Introduction

In spite of the well-known importance of wild flora as a source of food and medicinal substances, more studies on the diversity and agronomic value of these plant species are still needed (Arena and Vater, 2005). Areas with indigenous flora offer non-domesticated plants (Monge et al., 2000), like *Berberis* genus in Patagonia (Orsi, 1984) for these purposes. *Berberidaceae* in the widest sense is a small family, consisting of 10 to 12 genera and about 600 species, with many as 500 of these belonging to *Berberis* L., widely distributed in both the Old World and New World (Nowicke and Skvarla, 1981). In Patagonia, *Berberis* genus is well represented by 16 species of native shrubs and they are distributed from Neuquén to Tierra del Fuego (Arena and Curvetto, 2008). However, according to a later classification of the genus (Landrum, 1999), the number of species is less than previous studies cited by Orsi (1984), as Landrum groups the species *B. buxifolia*, *B. microphylla* and *B. heterophylla* under *B. microphylla* G. Forst, postulating that the differences among them may fluctuate to retain its range of species.

In particular, *B. microphylla* (ex *B. buxifolia* Lam.), commonly known as “calafate”, is an evergreen shrub that is present throughout the region mentioned, prevalent on the island of Tierra del Fuego, and over other species of *Berberis*. *B. microphylla* has growing economic potential due to the production of fruits as a non-timber forest product (Tacón Clavain, 2004). In fact, its dark blue berries are consumed fresh, as jams and preserves, and are used for the production of soft drinks and ice cream. Moreover, the fruits have a high content of phenols and antioxidants (Arena and Curvetto, 2008; Arena et al., 2012). Some characteristics of its phenological phases (Arena et al., 2013 a; Arena and Radice, 2014), fruit composition and production (Arena and Curvetto, 2008; Arena et al., 2003; 2011; 2013 b) have already been studied in natural populations of this species.

Characterization of pollen grains is an important step for programs of genetic resource conservation and improvement, complementing basic studies of biological data that characterize genotypes (de Castro Nunes et al., 2012). Selection on male game-
trophophytes (pollen) to alter the genetic constitution of the subsequent sporophytic generation has been suggested as an interesting tool in plant breeding programs (Hormaza and Herrero, 1992). Pollen competitive ability describes the reproductive success of a pollen grain and can therefore be considered as equivalent to pollen fitness (Sari-Gorla and Frova, 2005). The variability in pollen performance (size, viability and germination) among genotypes of a natural population could be interpreted as a survival strategy to maximize reproduction fitness (Tejaswini, 2002), while enabling a capacity of response to environmental changes (Hedly et al., 2005). Nevertheless, aspects related to fertility and reproductive organs have not yet been studied on <em>Berberis microphylla</em>.

The objective of this research was to describe the morphology and anatomy of pollen grains of <em>Berberis microphylla</em> G. Forst genotypes growing spontaneously on the island of Tierra del Fuego (Argentina), and evaluate their vitality and germination.

2. Materials and Methods

<em>Plant material</em>

Flowers (<em>n</em>=20) in phase E (before anthesis according to Arena et al., 2011) were collected from each <em>B. microphylla</em> genotype grown near Ushuaia city, Tierra del Fuego (54° 48’ SL, 68° 19’ WL and 30 m asl) (Table 1), in October 2013. The flowers were immediately placed in Petri dishes with wet paper at 5°C for viability and germination studies.

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<th>Genotype number</th>
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<em>Pollen grain description and size</em>

Equatorial and polar diameters of the pollen grains (<em>n</em>=50, randomly selected) were measured for each studied genotype using a Leica DM 2500 microscope. The average of the two parameters for each pollen grain was then calculated.

<em>Light microscopy</em>

Button flowers were dehydrated in an ethanol series and embedded in Spurr’s resin. Thin sections (75-90 nm thick) were stained with uranyl acetate and lead citrate.

<em>Scanning electron microscopy</em>

Button flowers were dehydrated in an ethanol series and a critical point drying technique was employed. Samples were sputter coated with 20 nm gold and observed with a Philips XL 30 SEM.

<em>Transmission electron microscopy</em>

Anthers were pre-fixed in 2.5% glutaraldehyde in phosphate buffer (pH 7.2) for 2 h and then post-fixed in <em>OsO</em><sub>4</sub> at 2°C in the same buffer for 3 h. They were then dehydrated in ethanol series and embedded in Spurr’s resin. Thin sections (75-90 nm thick) were made on a Sorval ultramicrotome, stained with uranyl acetate and lead citrate (O’Brien and Mc Cully, 1981). Sections were observed with a Jeol-Jem 1200 EXII TEM at 85.0 kv.

<em>Pollen grain viability</em>

Pollen grains were hydrated with sucrose solution (15%) and treated with fluorescein diacetate (10%) and propidium iodine (2%) (Greissl, 1989). The number of viable and not viable pollen grains was recorded under optic microscope, with a minimum of 300 pollen grains per genotype.

<em>Pollen grain germination</em>

Pollen grains were put on micro drops of a saline solution composed of 2 x 10<sup>−3</sup> M <em>H</em><sub>2</sub><em>Bo</em><sub>4</sub> and 6 x 10<sup>−3</sup> M Ca (<em>NO</em><sub>3</sub>)<sub>2</sub> added with sucrose 30% (Dafni, 1992). Micro drops were placed on the inside of the lid of a petri dish in which 3 ml of water were added in the base to create a humid chamber. Incubation was at 21±2°C. The number of germinated and aborted pollen grains was recorded under optic microscope 24 h after the test started and performed with anthers conserved for 1, 10 and 21 days at 5°C.

<em>Data analysis</em>

Measurements were analyzed by ANOVA and Tukey’s test and chi-square test was employed to evaluate pollen vitality.
3. Results

Pollen grain description and size

Flowers collected in phase E had five stamens; the anthers were not yet dehiscent (Fig. 1A) although the pollen grains were already mature (Fig. 1B, 2A-B). A mature pollen grain is formed of a vegetative cell with a dense cytoplasm with numerous starch grains (Fig. 1B). *B. microphylla* pollen grains are spherical with a psilate punctate surface interrupted by mild cracks (Fig. 2 A-B), resembling a tennis ball.

The pollen wall is formed by an exine and intine of considerable thickness (Fig. 3A). Transmission electron micrographs make it possible to identify an exine with two different layers, the ektxine and the endexine (Fig. 3A). Ektxine is nearly amorphous and not organized into typical foot layer, columellae, and tectum units. Conversely, ektxine appears as an external irregular cover with channels and small enclosed areas which are electron translucent (Fig. 3). Endexine has greater electro-density than ektxine. Immediately below this, the intine is present, at least four times thinner than the exine and with a very low electro-density. Pollen grain wall appears different on the apertures or pore zone (Fig. 3B). Ektxine is less structured and is represented by nodules above the endexine.

The pollen grain diameters varied significantly among genotypes, from 40 to 47.26 µm (p≤0.001). The maximum values were observed for genotype 107, which was significantly higher than genotypes 124, 126, 177, and 201 (Fig. 4). Pollen grains of genotypes 124 and 201 were significantly smaller than genotypes 107, 111, 123 and 202 (Fig. 4).

![Fig. 1 - Androecium and pollen grain of *B. microphylla* flower in phase E. A) view of androecium with anther (a) no dehiscent; B-C) microphotograph of cross-section of anther and mature pollen grain with starch grains (arrowhead). Bars: A = 1 mm; B = 10 µm; C = 100 µm.](image)

![Fig. 2 - SEM micrograph of mature pollen grain of *B. microphylla*. A) view of microsporangium with pollen grain; B) detail of a pollen grain. Bars: A = 10,000 nm; B = 50 µm.](image)

![Fig. 3 - TEM micrograph of pollen grain wall of *B. microphylla*. A) Detail of different parts of exine: endexine (end), ektxine (ekt) and intine (i); B) Detail of pore zone with ektxine fragmented (arrows). Barras = A-B = 1 µm.](image)

![Fig. 4 - Mean diameter of *B. microphylla* pollen grains of the studied genotypes.](image)
Vital pollen grains were bright green while non-vital ones stained red (Fig. 5). In this latter case, another category was evaluated, the sub-vital pollen grains, those that can germinate but it is uncertain whether they can be efficient in fertilization. Pollen viability of different genotypes gave very different results \((p \leq 0.001)\). For genotypes 111, 123, 124 and 202, values of vital pollen grains were above 70%, while for genotype 108 the value was 51.47% (Fig. 6). In coincidence, the 108 genotype shows a 36.80% of sub-vital pollen grains, value significantly greater than that observed for the 111, 123, 124, 126, and 202 genotypes.

**Fig. 6** - Viability of B. microphylla pollen grains from the assessed genotypes. Columns with different letters indicate significant differences between genotypes and date assessed \((\chi^2 p < 0.05)\).

**Fig. 7** - Germination percentage of B. microphylla pollen grains from the assessed genotypes. Columns with different letters indicate significant differences between genotypes and date assessed \((\text{Tukey} p < 0.05)\).

**Pollen grain germination**

The pollen grain germination of B. microphylla was significantly different among genotypes \((p \leq 0.001)\) and days of conservation \((p \leq 0.001)\), and the interaction between the two factors studied \((p \leq 0.001)\) was significant (Fig. 7). Germination of the pollen grain was at maximal level after one day of collection in most genotypes, except for genotype 123 which showed a maximum value after 21 days of storage, although without significant differences between 10 and 21 days (Fig. 7). Genotypes 124 and 125 showed percentages of pollen grain germination up to 70%, while genotype 201 presented a maximum value of 87.03% after one day of the collected flowers (Fig. 7). The pollen germination rate remained unchanged among the three tested dates, except for genotypes 122, 126 and 177. In addition, the values were significantly lower after 21 days of conservation for genotypes 122, 126 and 177 (Fig. 7). This decrease in the percentage of pollen germination is in accordance with the percentage of aborted pollen grains (data not shown). ANOVA values were 0.001 for both date and genotype factor and their interaction. Furthermore, the genotypes 108 and 202 showed an increase in the percentage of germinated pollen grains between the second and third test, i.e. the values obtained after 21 days of flower conservation were higher than those obtained with 10 days of storage but these differences were not significant (Fig. 7).

Finally, a significant negative correlation between the pollen grain germination and the pollen grain size was found \((r = -0.252; p \leq 0.001)\).

**4. Discussion and Conclusions**

It was observed that flower differentiation on B. microphylla started 12 weeks after bud break in coincidence with the end of the first fruit growth phase...
(Arena and Radice, 2014). Nevertheless, final development of male gametes occurs the following spring when the flower bud elongates (data not shown). Mature pollen grains can be found in the stage of lower emergence code 59 according to the BBCH scale (Arena et al., 2013 a). There are few published studies on the pollen of Berberis species. Erdtman (1952) and Heusser (1971) described the pollen grains of the Berberis genus and they determined that the pollen grains measured an average of 30 to 65 µm with an exine 2-3 µm thick. Nowicke and Skvarla (1981) emphasized the presence of irregular apertures, a psilate surface and an unstratified exine. In addition, the exine of B. micropyllyla shows an endexine as a prominent fibrous-granular layer and the ektexine with cavities and channels which suggest the endexine.

Pollen performance traits are often genetically based (Hedly et al., 2005; Hove and Mazer, 2013), however they could be also affected by differences in the nutritive status of the developing pollen grains as well as by the environmental conditions (Hedly et al., 2005). The size of pollen grains is considered to be one indicator of their viability (i.e. germinability and pollen tube growth rate), while the proportion of large pollen grains has been used to estimate pollen performance. Variation in pollen grain size among plants has been documented for several species (Varis et al., 2011). Larger pollen grains are thought to contain more resources for germination and, thus, have greater viability than smaller grains (Dufay et al., 2008). However, in B. microphylla a negative correlation between pollen size and germination percentage was observed, as was also found in Pinus sylvestris (Varis et al., 2011), as the growth of the pollen tube may be more dependent on pollen storage than pollen size. Good fertilization is directly related to very good pollen viability. It is estimated that the value of viability should be above 70% for fruit production (Urquieta, 2010). On the other hand, pollen grains which exceed 50% viability would be the only one that can be selected for use as male parents (Urquieta, 2010). All genotypes of B. microphylla tested presented viability values above 50%, in other words having very good prospects for fruit production. Urquieta (2010) found similar results when pollen grains of B. bidentata, B. darwinii, B. parodii and B. trigona were tested. Pollen grain viability of these species was variable between 59.6 and 74.1%.

It has been found that high percentages of pollen viability are due to high degrees of adaptability of the species to different environmental conditions (Kelly et al., 2002). In effect, high frequency of viable pollen reflects the adaptability of the species, since environmental plasticity submitted by pollen allows the genotype a satisfactory performance to different environmental conditions (Paupiére et al., 2014). On the other hand, pollen viability was influenced by relative humidity, temperature, atmospheric composition, and oxygen pressure after release from the anthers (Bots and Mariani, 2005), so it is expected that the experimental values obtained for B. microphylla are less than true values.

Pollen germination under in vitro conditions always produces lower values than those obtained with the viability test (Ontivero et al., 2006). In the present study, this premise is true for genotypes 107, 111, 123, 124, and 202 (Figs. 6 and 7). On the contrary, all other genotypes showed viability values higher than those obtained by germination viability test. Note that after 21 days of harvested flowers, pollen germination values obtained were only 10% lower than viability values for genotypes 108 and 123. These results could be due to the protective effect that antioxidants have on pollen. In effect, it is well known that secondary metabolites produced in the tapetum, such as phenolic compounds, can spread to the pollen and play a role in pollen colour, in the attraction of pollinators, in pollen tube germination, and in protection against abiotic stress of pollen (Paupiére et al., 2014). Pollen germination and tube growth is largely due to the presence of flavonols in mature pollen grains (Yistra et al., 1992). Although the content of flavonols was not measured in the anthers of B. microphylla, it is well known that flavonols are present in Berberis species (Končić et al., 2010). In effect, accumulation patterns of phenolic compounds during fruit growth and ripening in B. buxifolia (B. microphylla) was studied by Arena et al. (2012). It is likely that high values obtained from in vitro germination of pollen grains could be explained by the protective effect that these compounds perform.

The variability found in pollen performance (size, viability and germination) among genotypes of the natural population of B. microphylla, and its correlations, suggest the existence of pollen competition leading to unequal reproductive success in this species, as was observed for Camellia sinensis by Muoki et al. (2007).

The significant variability found in pollen performance (size, viability and germination) among B. microphylla genotypes from a natural population could be interpreted as a highlighted survival strategy
to maximize reproduction fitness, with a marked capacity of response to environmental changes. High viable pollen frequency, together with germination percentages observed in all the genotypes tested, could indicate a good fertilization process. The correlation observed between size and germination percentage could be used as a marker of pollen grain performance, with these findings representing the first antecedents useful for *B. microphylla* breeding.

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